



The Chicago Consensus on Peritoneal Surface Malignancies: Management of Appendiceal Neoplasms

Chicago Consensus Working Group

Chicago, IL

ABSTRACT The Chicago Consensus Working Group provides multidisciplinary recommendations for the management of appendiceal neoplasms specifically related to the management of peritoneal surface malignancies. These guidelines are developed with input from leading experts including surgical oncologists, medical oncologists, pathologists, radiologists, palliative care physicians, and pharmacists. These guidelines recognize and address the emerging need for increased awareness in the appropriate management of peritoneal surface disease. They are not intended to replace the quest for higher levels of evidence.

APPENDICEAL NEOPLASMS WITH PERITONEAL METASTASES

This article provides multidisciplinary recommendations for the management of peritoneal surface malignancies of appendiceal origin and constitutes 1 article in a series composed by the Chicago Consensus Working Group for the Management of Peritoneal Surface Malignancies.^{1–10} Information regarding formation of the Chicago Consensus Group and explanation of the working group's consensus methodology is discussed.^{1,2}

The peritoneum is the most common site of metastasis of appendiceal neoplasms. Peritoneal metastases frequently occur as synchronous metastases but can also be seen as metachronous metastases. The incidence of peritoneal metastases varies depending on the histologic characteristics of the primary appendiceal neoplasm and the presence of perforation and has been reported to range from 5 to > 50%.¹¹

In the initial evaluation of these patients, the histologic subtype of the tumor, the burden of disease on the peritoneal surfaces, and the presence of nonperitoneal metastatic sites must be assessed. A thorough evaluation of the resected primary tumor and/or biopsy specimens of peritoneal metastatic sites should be performed by an experienced pathologist. Cross-sectional imaging with contrast-enhanced computed tomography (CT) or magnetic resonance imaging is the preferred imaging modality to evaluate the peritoneal burden of disease, although significant limitations exist in the ability of any imaging modality to detect small-volume disease. Additional staging imaging such as chest CT is recommended for staging of advanced and/or recurrent low-grade mucinous tumors and for high-grade tumors, including goblet cell adenocarcinoma (GCA). Positron emission tomography is not recommended for the evaluation of mucinous tumors. Levels of CEA, CRP, Ca 19-9, CA-125, and chromogranin A (for GCA only) are often elevated, and their measurement can be useful in patients with peritoneal metastases from appendiceal neoplasms.^{12–18} See Figs. 1, 2 and 3, management pathways for goblet cell adenocarcinoma, low-grade appendiceal mucinous neoplasm, and appendiceal adenocarcinoma.

Principles of Pathology

The histologic evaluation of a peritoneal biopsy specimen and/or the primary tumor provides the basis for determining the specific subtype of appendiceal neoplasm

The collaborators for the Chicago Consensus Working Group are listed in the acknowledgments.

Correspondence to: K. Turaga, MD
Department of Surgery, University of Chicago, Chicago, IL
e-mail: kturaga@surgery.bsd.uchicago.edu

© American Cancer Society and Society of Surgical Oncology 2020

First Received: 3 July 2019;
Published Online: 13 April 2020

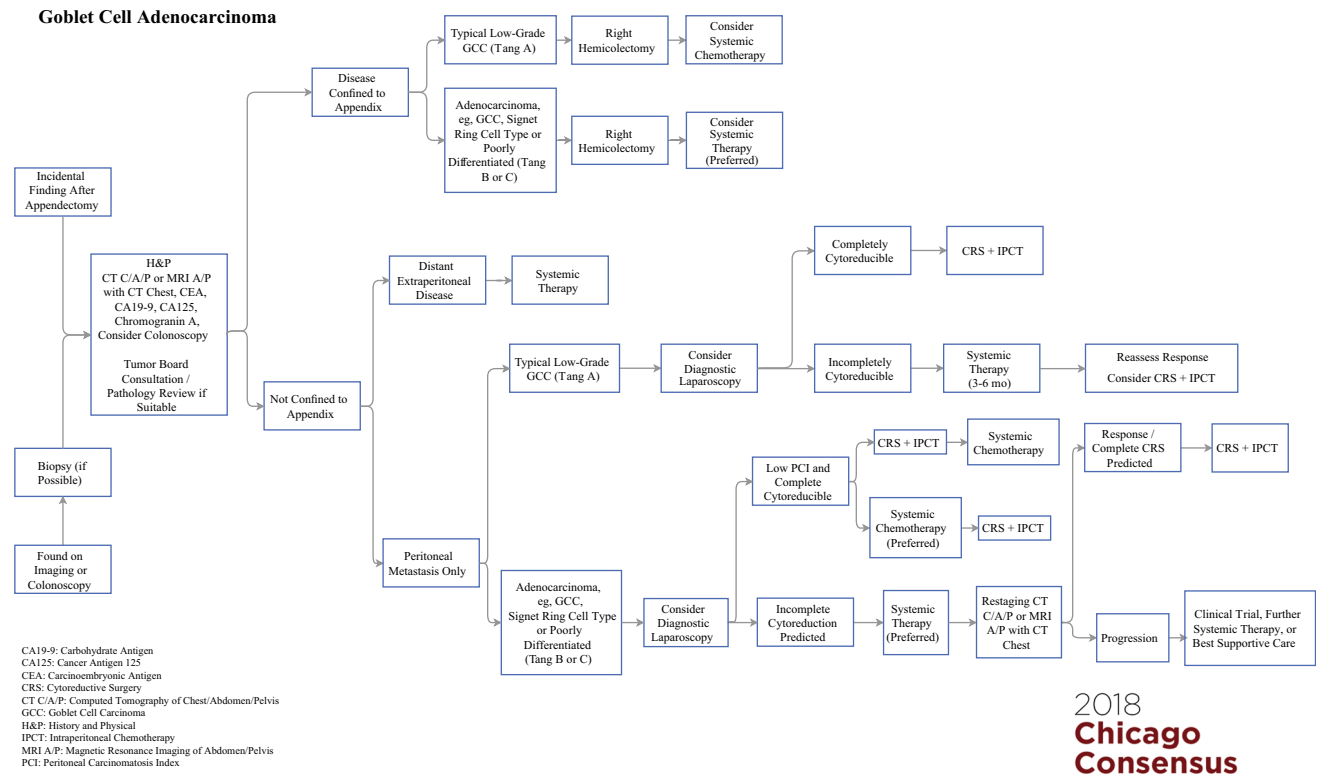


FIG. 1 Goblet cell adenocarcinoma management pathway

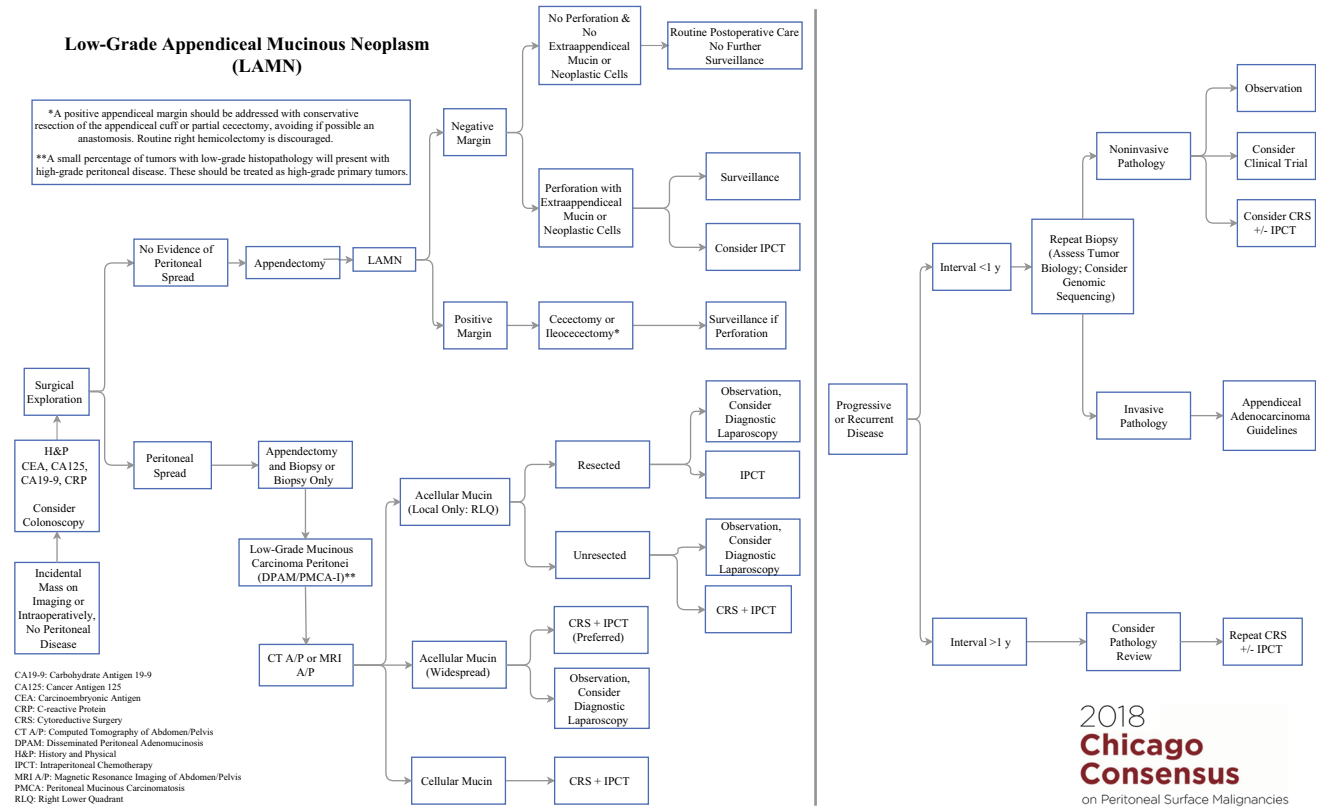


FIG. 2 Low-grade appendiceal mucinous neoplasm management pathway

Appendiceal Adenocarcinoma

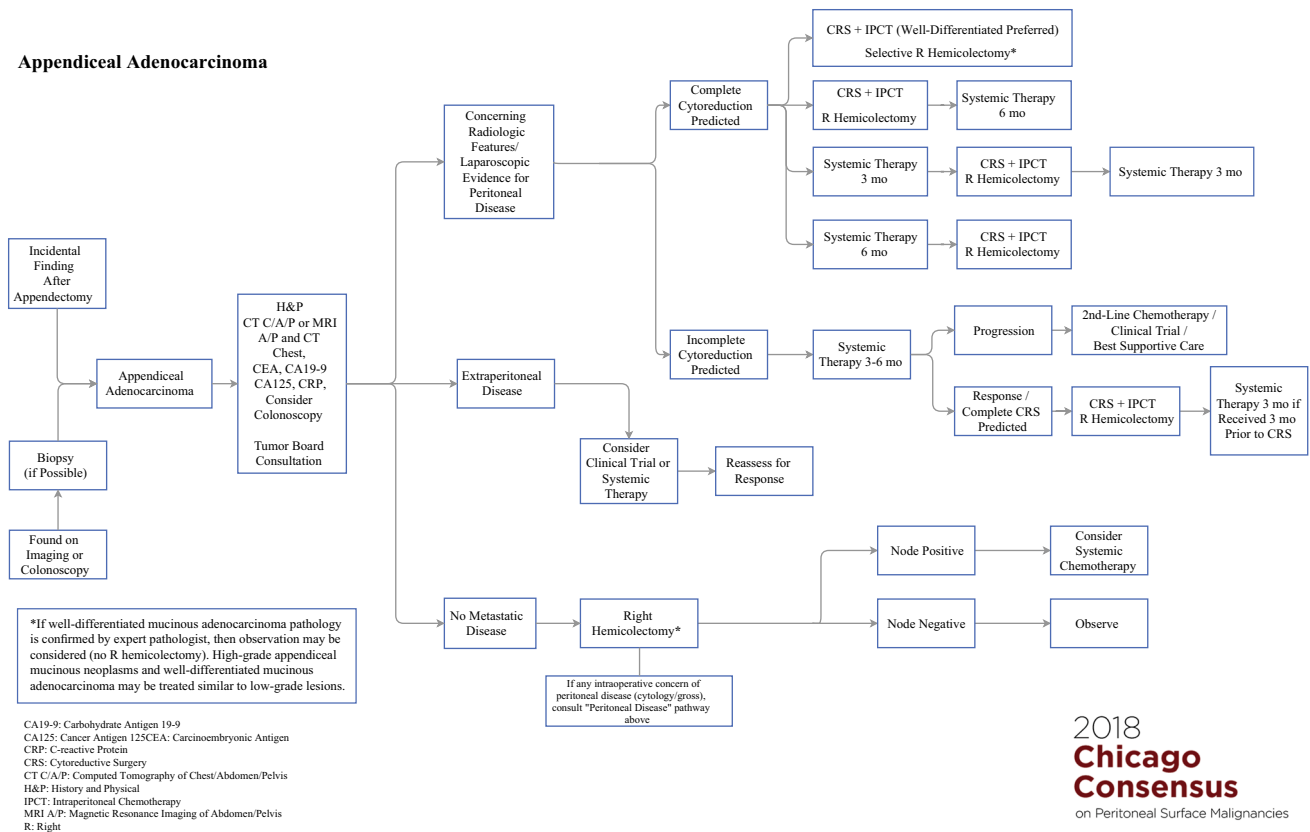


FIG. 3 Appendiceal adenocarcinoma management pathway

and its grade. Standard hematoxylin–eosin staining and immunohistochemistry methods are the basis of the pathological evaluation of appendiceal tumors. However, because of controversial and confusing nomenclature of several appendiceal neoplastic entities, the pathological assessment should be performed by a pathologist experienced in appendiceal and peritoneal tumors.^{11,19}

On gross evaluation, most appendices with primary mucinous neoplasia show appendiceal dilatation as a result of abnormal accumulation of mucin within the lumen. The gross and macroscopic examination of these neoplasms should include the location of the tumor within the appendix (tip vs body vs base), the size of the tumor, the distance of tumor from the proximal resection margin (when possible), the presence or absence of mucin deposits on the serosa, the presence or absence of perforation, and the presence of any solid areas. Following formalin fixation, the entire appendix should be submitted for histologic examination, with the margin of resection separately designated.

After formalin fixation, peritoneal tissue should be measured and examined for mucin deposits and any firm nodules. If the specimen consists solely of mucin or contains a separate mucinous component, a measurement should also be given for this component and careful

inspection for solid areas should be performed. For each specimen part, it is reasonable to submit 1 section per centimeter of mucinous deposit initially. If initial sections demonstrate high-grade disease, then there is no added value to additional tissue sections. However, detection of only acellular mucin or low-grade disease should prompt additional tissue sections, including submission of all mucinous material when feasible. Because of the low cellularity in many of these specimens, intraoperative assessment via frozen section is not recommended. Mucinous specimens can be processed and embedded in paraffin according to standard laboratory protocols. If molecular analysis is needed, microdissection may be required to obtain sufficient tumor DNA.

Biopsies of the peritoneal tumor deposits should be performed using image-guided core-needle technique or via a diagnostic laparoscopy. Positioning of the needle tracts and trocars should take into consideration the potential risk of seeding along the tract (midline preferred).

The presence of mucin without epithelial cells in a peritoneal biopsy specimen should not be classified as benign disease or a nondiagnostic sample; its significance and further management decisions should be determined by providers experienced in the management of appendiceal tumors. Cytologic evaluation of mucinous ascites can

identify glandular cells, although low tumor cellularity and lack of consensus guidelines limit the value of this method.

The pathological evaluation of the primary appendiceal tumor specimen should include the following features:

- Organ and site.
- Primary diagnosis according to *Digestive System Tumours*, volume 1 of *WHO Classification of Tumours*, fifth edition.^{20–22} For GCA, histologic classification as typical GCA (Tang group A), signet ring cell type (Tang group B), or poorly differentiated carcinoma type (Tang group C) should be noted.²³
- Histologic grade, specifically including the presence of signet ring cells.
- Tumor stage.
- Presence of perforation of appendix or tumor.
- Presence of mucin or neoplastic epithelial cells on the appendiceal serosa or mesoappendix.
- Presence of lymphatic vessel invasion, large vessel invasion, or perineural invasion.
- Status of margins.
- Status of lymph nodes.

The pathological evaluation of the peritoneal metastases should include the following:

- Histologic type according to the World Health Organization classification.^{20–22}
- Histologic grade.
- Presence of signet ring cells, including percentage of tumor composed of signet ring cells.
- Results of ancillary studies, including immunohistochemistry to determine the potential origin of the tumor.

Principles of Systemic Chemotherapy

Limited evidence regarding the role of systemic chemotherapy in appendiceal neoplasms is available, and the existing evidence shows limited efficacy for the most common subtypes. Nonmucinous epithelial appendiceal neoplasms are generally approached and treated using colorectal cancer treatment pathways (Fig. 3).

Low-grade mucinous appendiceal neoplasms and low-grade mucinous carcinoma peritonei of appendiceal origin are generally considered indolent and show minimal to no responsiveness to systemic chemotherapy (Fig. 2).^{24–26} Therefore, patients who are candidates for surgical cytoreduction are treated with surgery and hyperthermic intraperitoneal chemotherapy (HIPEC) followed by surveillance. For patients with unresectable disease and/or recurrent disease not amenable to resection, systemic therapy can be used according to the treatment algorithms for mucinous colorectal cancer (Fig. 2).^{27,28}

For patients with mucinous appendiceal adenocarcinoma, nonmucinous appendiceal adenocarcinoma, signet ring cell adenocarcinoma, high-grade mucinous carcinoma peritonei, or high-grade and/or advanced-stage GCA, systemic chemotherapy is used as part of multimodality therapy akin to the treatment of colorectal cancer.²⁹ There is limited evidence regarding efficacy in appendiceal cancer specifically, but several studies suggest similar response rates in these cancer subtypes.^{27,29} It is worth noting that not all patients with GCA necessarily require systemic chemotherapy, especially those with Tang group A tumors (Fig. 1). However, limited evidence to help guide this decision is available.

For patients who are candidates for cytoreductive surgery and HIPEC, systemic chemotherapy can be used preoperatively, postoperatively, or both.^{24,26,30} Following standards used in other colorectal cancer settings, a combined total of 6 months of chemotherapy is usually recommended.

Principles of Surgery

Surgery with a curative intent can be considered for patients with either low-grade or high-grade subtypes of appendiceal neoplasms and peritoneal metastases. However, inclusion criteria and expected outcomes are notably different.

Peritoneal metastases of appendiceal origin are often detected incidentally at the time of appendectomy or diagnostic laparoscopy. In these situations surgical resection should be limited to steps that will help establish an accurate diagnosis (most commonly peritoneal biopsy and appendectomy if feasible). More extensive resections or partial debulking should be avoided at this time.

Complete cytoreduction is the main goal of surgical management of peritoneal metastases and is the most basic principle in preoperative decision-making. It is important to note that for low-grade appendiceal mucinous neoplasms and well-differentiated mucinous adenocarcinoma, right hemicolectomy is not always necessary.^{31,32}

Cross-sectional imaging with or without diagnostic laparoscopy is used to estimate the likelihood of complete cytoreduction. Multifocal bowel obstruction, extensive involvement of the small bowel, biliary obstruction, and ureteral obstruction (especially bilateral) usually preclude successful curative-intent surgery.

For patients with low-grade mucinous neoplasm (grade 1), cytoreductive surgery with HIPEC is the treatment of choice. Excellent long-term survival can be achieved even in patients with a high burden of disease if complete cytoreduction can be achieved.^{33–35} Patients who develop peritoneal recurrence after initial cytoreduction may be candidates for repeat surgical procedures.^{36–38}

Patients with adenocarcinoma and peritoneal metastases are less likely than those with low-grade primary neoplasms and peritoneal disease to have complete cytoreduction. These patients also have worse survival rates when a high disease burden is present.^{39,40} Preoperative predictive scoring systems can be helpful in preoperative decision-making,⁴¹ although an absolute cut-off value of peritoneal disease burden has not been established.

Surgeons performing cytoreductive surgical procedures must be expert in assessing all of the peritoneal surfaces and knowledgeable about the typical patterns of involvement with different types of peritoneal surface malignancies. They need to have expertise performing multiple visceral resections and knowledge of standard peritonectomy techniques. Palliative debulking procedures (or incomplete cytoreductions/ablations) are generally discouraged except for patients with low-grade tumors; these procedures can be considered in appropriately selected patients. The burden of disease must be documented with a standard scoring system such as the peritoneal cancer index (PCI) score, and completion of cytoreduction should be documented with a CC or R score.⁴²

Principles of Intraperitoneal Chemotherapy

Intraperitoneal chemotherapy has been applied in most studies that assessed complete cytoreduction for peritoneal metastases of appendiceal origin. The only randomized controlled study on this subject was designed to assess outcomes of cytoreductive surgery (CRS) and HIPEC in patients with colorectal cancer but included some patients with appendiceal adenocarcinoma. This study showed improved survival for patients treated with CRS and HIPEC plus systemic chemotherapy as compared with systemic chemotherapy with or without palliative surgery.⁴³ The first and only randomized trial of patients with appendiceal cancer was recently reported. The primary aim of this trial was to evaluate the toxicity of HIPEC with mitomycin C versus HIPEC with oxaliplatin. Progression-free and overall survival did not differ between groups, but the overall results confirmed that long-term survival is possible if complete cytoreduction is achieved.⁴⁴

Large cohort studies of complete cytoreduction using peritonectomy procedures and visceral resection without HIPEC or early postoperative intraperitoneal chemotherapy are not available.

Current regimens that may be used for intraperitoneal chemotherapy are as follows:

- Mitomycin 30 mg at time 0, and 10 mg at time 60 for 90 min.

- Mitomycin 30 mg/m² for 90 to 120 min.
- Mitomycin 15 mg/m² plus doxorubicin 15 mg/m² for 90 min.
- Oxaliplatin 300 mg/m² for 30 min.

ACKNOWLEDGMENT Collaborators: Darryl Schuitevoerder, MBBS, University of Chicago, Department of Surgery, Chicago, IL; Alejandro Plana, BA, University of Chicago, Department of Surgery, Chicago, IL; Francisco J. Izquierdo, MD, Clínica Santa María, Department of Surgery, Providencia, Chile; Konstantinos I. Votanopoulos, MD, PhD, Wake Forest, Surgery, Winston-Salem, NC; James C. Cusack Jr, MD, Massachusetts General Hospital, Department of Surgery, Boston, MA; Lana Bijelic, MD, Inova Schar Cancer Institute, Surgical Oncology, Fairfax, VA; Clifford S. Cho, MD, University of Michigan Medical School, Department of Surgery, Ann Arbor, MI; Alexandra Gangi, MD, Cedars-Sinai Medical Center, Surgical Oncology, Department of Surgery, Los Angeles, CA; Erin W. Gilbert, MD, Oregon Health & Science University, Department of Surgery, Portland, OR; Martin D. Goodman, MD, Tufts Medical Center, Surgical Oncology, Boston, MA; Anand Govindarajan, MD, MSc, Sinai Health System, University of Toronto, Division of General Surgery, Toronto, ON, Canada; Vadim Gushchin, MD, Institute for Cancer Care at Mercy, Surgical Oncology, Baltimore, MD; Chukwuemeka Ihemelandu, MD, MedStar Georgetown University Medical Center, Surgical Oncology, Washington, DC; Kaitlyn J. Kelly, MD, University of California San Diego, Surgery, La Jolla, CA; Ryan P. Merkow, MD, MS, Northwestern University, Feinberg School of Medicine, Department of Surgery, Evanston, IL; Sam G. Pappas, MD, Rush University Medical Center, Surgery, Chicago, IL; Daniel E. Abbott, MD, University of Wisconsin, Department of Surgery, Madison, WI; Steven A. Ahrendt, MD, University of Colorado, Department of Surgery, Aurora, CO; Mazin Al-kasspoles, MD, University of Kansas Medical Center, Department of Surgery, Division of Surgical Oncology, Kansas City, KS; Lindsay Alpert, MD, University of Chicago, Department of Pathology, Chicago, IL; Farin Amersi, MD, Cedars-Sinai Medical Center, Los Angeles, CA; Amanda K. Arrington, MD, University of Arizona, Surgery, Tucson, AZ; Brian Badgwell, MD, MS, MD Anderson Cancer Center, Surgical Oncology, Houston, TX; Robert M. Barone, MD, MS, Sharp HealthCare, UCSD School Medicine Division Surgical Oncology, Surgery, La Jolla, CA; Joel M. Baumgartner, MD, MAS, University of California San Diego, Surgery, La Jolla, CA; Dan G. Blazer III, MD, Duke University, Surgery, Durham, NC; Richard N. Berri, MD, Ascension St. John Hospital, Surgical Oncology, Grosse Pointe Woods, MI; Charles Komen Brown, MD, PhD, AMITA Health, Surgery, La Grange, IL; Daniel V. Catenacci, MD, University of Chicago, Medicine, Chicago, IL; Carlos H. F. Chan, MD, PhD, University of Iowa Hospitals and Clinics, Department of Surgery, Iowa City, IA; M. Haroon A. Choudry, MD, University of Pittsburgh, Surgery, Surgical Oncology, Pittsburgh, PA; Callisia N. Clarke, MD, MS, Medical College of Wisconsin, Surgery, Milwaukee, WI; Jordan M. Cloyd, MD, The Ohio State University, Department of Surgery, Columbus, OH; Abraham H. Dachman, MD, UChicago Medicine, Radiology, Chicago, IL; Jeremiah L. Deneve, DO, University of Tennessee Health Science Center, Surgery, Memphis, TN; Sean P. Dineen, MD, Moffitt Cancer Center, GI Oncology, Tampa, FL; Leopoldo J. Fernandez, MD, Virginia Commonwealth University Massey Cancer Center, Hunter Holmes McGuire VAMC, Surgical Oncology, Richmond, VA; Oliver S. Eng, MD, University of Chicago, Surgery, Chicago, IL; James W. Fleshman, Jr, MD, Baylor University Medical Center, Department of Surgery, Dallas, TX; T. Clark Gamblin, MD, MS, MBA, Medical College of Wisconsin, Department of Surgery, Milwaukee, WI;

Georgios V. Georgakis, MD, PhD, Stony Brook University, Department of Surgery, Division of Surgical Oncology, Stony Brook, NY; **Travis E. Grotz, MD**, Mayo Clinic, Department of Surgery, Rochester, MN; **Nader Hanna, MD**, University of Maryland School of Medicine, Surgery, Baltimore, MD; **Carla Harmath, MD**, University of Chicago, Radiology, Chicago, IL; **John Hart, MD**, University of Chicago, Department of Pathology, Chicago, IL; **Andrea Hayes-Jordan, MD**, University of North Carolina School of Medicine, Department of Surgery, Division of Pediatric Surgery, Chapel Hill, NC; **Aliya N. Husain, MD**, University of Chicago, Pathology, Chicago, IL; **Kamran Idrees, MD, MSCI, MMHC**, Vanderbilt University Medical Center, Department of Surgery, Nashville, TN; **Haejin In, MD, MBA, MPH**, Montefiore Medical Center, Surgery, Bronx, NY; **David Jiang, MD**, University of Chicago, Department of Surgery, Chicago, IL; **John M. Kane III, MD**, Roswell Park Comprehensive Cancer Center, Surgical Oncology, Buffalo, NY; **Timothy J. Kennedy, MD, MBA**, Rutgers Cancer Institute of New Jersey, GI Surgical Oncology, New Brunswick, NJ; **Xavier M. Keutgen, MD**, University of Chicago, Surgery, Division of Surgical Oncology, Endocrine Surgery Research Program, Chicago, IL; **Hedy Lee Kindler, MD**, University of Chicago, Section of Hematology/Oncology, Chicago, IL; **Byrne Lee, MD**, City of Hope, Division of Surgical Oncology, Duarte, CA; **Chih-Yi Liao, MD, BA**, University of Chicago, Section of Hematology/Oncology, Department of Medicine, Chicago, IL; **Ugwuji N. Maduekwe, MD, MSc**, The University of North Carolina at Chapel Hill, Department of Surgery, Division of Surgical Oncology and Endocrine Surgery, Chapel Hill, NC; **Grace Z. Mak, MD**, University of Chicago, Department of Surgery, Section of Pediatric Surgery, Chicago, IL; **Lloyd A. Mack, MD, MSc**, University of Calgary, Departments of Surgery and Oncology, Calgary, AB, Canada; **Melvy Sarah Mathew, MD**, University of Chicago, Department of Radiology, Chicago, IL; **Marcovalerio Melis, MD**, NYU School of Medicine, Surgery, New York, NY; **Nelya Melnitchouk, MD, MSc**, Brigham and Women's Hospital, Harvard Medical School, Department of Surgery, Boston, MA; **Joseph Misdraji, MD**, Massachusetts General Hospital, Department of Pathology, Boston, MA; **Harveshp Mogal, MD, MS**, Medical College of Wisconsin, Surgery, Milwaukee, WI; **Mecker G. Möller, MD**, University of Miami Miller School of Medicine, Department of Surgery, Division of Surgical Oncology, Miami, FL; **Garrett M. Nash, MD, MPH**, Memorial Sloan Kettering, Surgery, New York, NY; **Aytekin Oto, MD, MBA**, University of Chicago, Radiology, Chicago, IL; **Reetesh K. Pai, MD**, University of Pittsburgh, Department of Pathology, Pittsburgh, PA; **Colette R. Pameijer, MD**, Penn State College of Medicine, Department of Surgery, Hershey, PA; **Patricio M. Polanco, MD**, University of Texas Southwestern Medical Center, Department of Surgery, Division of Surgical Oncology, Dallas, TX; **Blase N. Polite, MD, MPP**, University of Chicago, Department of Medicine, Chicago, IL; **Sanjay S. Reddy, MD**, Fox Chase Cancer Center, Department of Surgery, Philadelphia, PA; **Richard Royal, MD**, MD Anderson Cancer Center, Surgical Oncology, Houston, TX; **David P. Ryan, MD**, Massachusetts General Hospital, MGH Cancer Center, Boston, MA; **George Salti, MD**, Edward-Elmhurst Health and University of Illinois at Chicago, Surgical Oncology, Chicago, IL; **Armando Sardi, MD**, Mercy Medical Center, Surgical Oncology, Baltimore, MD; **Maheswari Senthil, MD**, Loma Linda University Health, Surgical Oncology, Loma Linda, CA; **Namrata Setia, MD**, University of Chicago, Department of Pathology, Chicago, IL; **Scott K. Sherman, MD**, University of Chicago, Surgery, Chicago, IL; **Lucas Sideris, MD, FRCSC**, University of Montreal, Surgery, Montreal, QC, Canada; **Joseph Skitzki, MD**, Roswell Park Comprehensive Cancer Center, Department of Surgical Oncology, Buffalo, NY; **Jula Veerapong, MD**, University of California San Diego, Surgical Oncology, La Jolla, CA; **Michael G. White, MD, MSc**, University of Chicago, Department of Surgery, Chicago, IL; **Joshua H. Winer, MD**, Emory

University, Division of Surgical Oncology, Atlanta, GA; **Shu-Yuan Xiao, MD**, University of Chicago, Chicago, IL, and Wuhan University Zhongnan Hospital, Department of Pathology, Wuhan, China; **Rhonda K. Yantiss, MD**, Weill Cornell Medicine, Pathology and Laboratory Medicine, New York, NY; **Nita Ahuja, MD, MBA**, Yale University, Surgery, New Haven, CT; **Wilbur Bowne MD**, Drexel University College of Medicine, Department of Surgery, Philadelphia, PA; **Andrew M. Lowy, MD**, UC San Diego Health, Department of Surgery, La Jolla, CA; **H. Richard Alexander Jr, MD**, Rutgers Cancer Institute of New Jersey, Division of Surgical Oncology, New Brunswick, NJ; **Jesus Esquivel, MD**, Frederick Memorial Hospital, Surgical Oncology, Frederick, MD; **Jason M. Foster, MD**, University of Nebraska/Nebraska Medicine, Surgery, Division of Surgical Oncology, Omaha, NE; **Daniel M. Labow, MD**, Icahn School of Medicine at Mount Sinai, Department of Surgery, New York, NY; **Laura A. Lambert, MD**, Huntsman Cancer Institute/University of Utah, General Surgery, Salt Lake City, UT; **Edward A. Levine, MD**, Wake Forest University, Surgical Oncology, Winston-Salem, NC; **Charles Staley, MD**, Emory University School of Medicine, Department of Surgery, Atlanta, GA; **Paul H. Sugarbaker, MD**, MedStar Washington Hospital Center, Washington, DC; **David L. Bartlett, MD**, University of Pittsburgh, Surgery, Pittsburgh, PA; **Kiran Turaga, MD, MPH**, University of Chicago, Department of Surgery, Chicago, IL.

FUNDING The Irving Harris Foundation and the University of Chicago.

DISCLOSURES James C. Cusack reports grants from Lumicell Inc. outside the submitted work. Carla Harmath serves on the medical advisory council of Accumen. Hedy Kindler reports personal fees and non-financial support from Inventiva, AstraZeneca, Boehringer Ingelheim, Merck, and Pareto; personal fees from Aldeyra Therapeutics, Bayer, BMS, Erytech, Five Prime Therapeutics, Ipsen Pharmaceuticals, Kyowa, and MedImmune; and funds to support clinical trials at her institution from Aduro, AstraZeneca, Bayer, BMS, Deciphera, GSK, Lilly, Merck, MedImmune, Polaris, Verastem, and Blueprint, all outside the submitted work. Chih-Yi Liao reports personal fees from Eisai, Exelixis, and Klus Pharma outside the submitted work. Garrett M. Nash reports non-financial support from Intuitive outside the submitted work. Aytekin Oto reports grants from Philips Healthcare, Guerbet, and Profound Healthcare, and serves as a medical advisory board member for Profound Healthcare, all outside the submitted work. David P. Ryan reports personal fees from and equity in MPM Capital, equity in Acworth Pharmaceuticals, personal fees from Oncorus, Gritstone Oncology, Maverick Therapeutics, TCR² Therapeutics, UpToDate, McGraw-Hill, and Johns Hopkins University Press, all outside the submitted work. Nita Ahuja reports grant funding from Cepheid and Astex, has served as a consultant to Ethicon, and has licensed methylation biomarkers to Cepheid. Jesus Esquivel reports personal fees from Eight Medical. The remaining authors disclosed no conflicts of interest.

REFERENCES

- Chicago Consensus Working Group. The Chicago Consensus Guidelines for Peritoneal Surface Malignancies: Introduction. *Ann Surg Oncol*. 2020. <https://doi.org/10.1245/s10434-020-08318-8>.
- Chicago Consensus Working Group. The Chicago Consensus on Peritoneal Surface Malignancies: Methodology. *Ann Surg Oncol*. 2020. <https://doi.org/10.1245/s10434-020-08317-9>.

3. Chicago Consensus Working Group. The Chicago Consensus on Peritoneal Surface Malignancies: Standards. *Ann Surg Oncol*. 2020. <https://doi.org/10.1245/s10434-020-08325-9>.
4. Chicago Consensus Working Group. The Chicago Consensus on Peritoneal Surface Malignancies: Management of Colorectal Metastases. *Ann Surg Oncol*. 2020. <https://doi.org/10.1245/s10434-020-08315-x>.
5. Chicago Consensus Working Group. The Chicago Consensus on Peritoneal Surface Malignancies: Management of Gastric Metastases. *Ann Surg Oncol*. 2020. <https://doi.org/10.1245/s10434-020-08320-0>.
6. Chicago Consensus Working Group. The Chicago Consensus on Peritoneal Surface Malignancies: Management of Peritoneal Mesothelioma. *Ann Surg Oncol*. 2020. <https://doi.org/10.1245/s10434-020-08324-w>.
7. Chicago Consensus Working Group. The Chicago Consensus on Peritoneal Surface Malignancies: Management of Ovarian Neoplasms. *Ann Surg Oncol*. 2020. <https://doi.org/10.1245/s10434-020-08322-y>.
8. Chicago Consensus Working Group. The Chicago Consensus on Peritoneal Surface Malignancies: Management of Neuroendocrine Tumors. *Ann Surg Oncol*. 2020. <https://doi.org/10.1245/s10434-020-08321-z>.
9. Chicago Consensus Working Group. The Chicago Consensus on Peritoneal Surface Malignancies: Management of Desmoplastic Small Round Cell Tumor, Breast, and Gastrointestinal Stromal Tumor. *Ann Surg Oncol*. 2020. <https://doi.org/10.1245/s10434-020-08319-7>.
10. Chicago Consensus Working Group. The Chicago Consensus on Peritoneal Surface Malignancies: Palliative Care Considerations. *Ann Surg Oncol*. 2020. <https://doi.org/10.1245/s10434-020-08323-x>.
11. Carr NJ, Bibeau F, Bradley RF, et al. The histopathological classification, diagnosis and differential diagnosis of mucinous appendiceal neoplasms, appendiceal adenocarcinomas and pseudomyxoma peritonei. *Histopathology*. 2017;71(6):847–858.
12. Carmignani CP, Hampton R, Sugarbaker CE, Chang D, Sugarbaker PH. Utility of CEA and CA 19-9 tumor markers in diagnosis and prognostic assessment of mucinous epithelial cancers of the appendix. *J Surg Oncol*. 2004;87(4):162–166.
13. Koh JL, Liauw W, Chua T, Morris DL. Carbohydrate antigen 19-9 (CA 19-9) is an independent prognostic indicator in pseudomyxoma peritonei post cytoreductive surgery and perioperative intraperitoneal chemotherapy. *J Gastrointest Oncol*. 2013;4(2):173–181.
14. Wagner PL, Austin F, Sathiah M, et al. Significance of serum tumor marker levels in peritoneal carcinomatosis of appendiceal origin. *Ann Surg Oncol*. 2013;20(2):506–514.
15. Chua TC, Chong CH, Liauw W, Zhao J, Morris DL. Inflammatory markers in blood and serum tumor markers predict survival in patients with epithelial appendiceal neoplasms undergoing surgical cytoreduction and intraperitoneal chemotherapy. *Ann Surg*. 2012;256(2):342–349.
16. Lohani K, Shetty S, Sharma P, Govindarajan V, Thomas P, Loggie B. Pseudomyxoma peritonei: inflammatory responses in the peritoneal microenvironment. *Ann Surg Oncol*. 2014;21(5):1441–1447.
17. Olsen IH, Holt N, Langer SW, et al. Goblet cell carcinoids: characteristics of a Danish cohort of 83 patients. *PLoS One*. 2015;10(2):e0117627.
18. Lamarca A, Nonaka D, Lopez Escola C, et al. Appendiceal goblet cell carcinoids: management considerations from a reference peritoneal tumour service centre and ENETS Centre of Excellence. *Neuroendocrinology*. 2016;103(5):500–517.
19. Carr NJ, Cecil TD, Mohamed F, et al., Peritoneal Surface Oncology Group International. A consensus for classification and pathologic reporting of pseudomyxoma peritonei and associated appendiceal neoplasia: the results of the Peritoneal Surface Oncology Group International (PSOGI) modified Delphi process. *Am J Surg Pathol*. 2016;40(1):14–26.
20. Misdraji J, Carr N, Pai R. Tumours of the appendix: mucinous neoplasm. In: WHO Classification of Tumours Editorial Board, editor. Digestive system tumours, vol 1, 5th ed. Lyon: International Agency for Research on Cancer; 2019. p. 144–146.
21. Misdraji J, Carr N, Pai R. Tumours of the appendix: adenocarcinoma. In: WHO Classification of Tumours Editorial Board, editor. Digestive system tumours, vol 1, 5th ed. Lyon: International Agency for Research on Cancer; 2019. p. 147–148.
22. Misdraji J, Carr NJ, Pai RK. Tumours of the appendix: goblet cell adenocarcinoma. In: WHO Classification of Tumours Editorial Board, editor. Digestive system tumours, vol 1, 5th ed. Lyon, France: International Agency for Research on Cancer; 2019. p. 149–151.
23. Tang LH, Shia J, Soslow RA, et al. Pathologic classification and clinical behavior of the spectrum of goblet cell carcinoid tumors of the appendix. *Am J Surg Pathol*. 2008;32(10):1429–1443.
24. Sugarbaker PH, Bijelic L, Chang D, Yoo D. Neoadjuvant FOLFOX chemotherapy in 34 consecutive patients with mucinous peritoneal carcinomatosis of appendiceal origin. *J Surg Oncol*. 2010;102(6):576–581.
25. Asare EA, Compton CC, Hanna NN, et al. The impact of stage, grade, and mucinous histology on the efficacy of systemic chemotherapy in adenocarcinomas of the appendix: analysis of the National Cancer Data Base. *Cancer*. 2016;122(2):213–221.
26. Blackham AU, Swett K, Eng C, et al. Perioperative systemic chemotherapy for appendiceal mucinous carcinoma peritonei treated with cytoreductive surgery and hyperthermic intraperitoneal chemotherapy. *J Surg Oncol*. 2014;109(7):740–745.
27. Shapiro JF, Chase JL, Wolff RA, et al. Modern systemic chemotherapy in surgically unresectable neoplasms of appendiceal origin: a single-institution experience. *Cancer*. 2010;116(2):316–322.
28. Farquharson AL, Pranesh N, Witham G, et al. A phase II study evaluating the use of concurrent mitomycin C and capecitabine in patients with advanced unresectable pseudomyxoma peritonei. *Br J Cancer*. 2008;99(4):591–596.
29. Lieu CH, Lambert LA, Wolff RA, et al. Systemic chemotherapy and surgical cytoreduction for poorly differentiated and signet ring cell adenocarcinomas of the appendix. *Ann Oncol*. 2012;23(3):652–658.
30. Milovanov V, Sardi A, Ledakis P, et al. Systemic chemotherapy (SC) before cytoreductive surgery and hyperthermic intraperitoneal chemotherapy (CRS/HIPEC) in patients with peritoneal mucinous carcinomatosis of appendiceal origin (PMCA). *Eur J Surg Oncol*. 2015;41(5):707–712.
31. Turaga KK, Pappas S, Gamblin TC. Right hemicolectomy for mucinous adenocarcinoma of the appendix: just right or too much? *Ann Surg Oncol*. 2013;20(4):1063–1067.
32. Votanopoulos KI, Shen P, Skardal A, Levine EA. Peritoneal metastases from appendiceal cancer. *Surg Oncol Clin N Am*. 2018;27(3):551–561.
33. Moran B, Baratti D, Yan TD, Kusamura S, Deraco M. Consensus statement on the loco-regional treatment of appendiceal mucinous neoplasms with peritoneal dissemination (pseudomyxoma peritonei). *J Surg Oncol*. 2008;98(4):277–282.
34. Esquivel J, Averbach A. Laparoscopic cytoreductive surgery and HIPEC in patients with limited pseudomyxoma peritonei of appendiceal origin. *Gastroenterol Res Pract*. 2012;2012:981245.
35. Smeenk RM, Verwaal VJ, Antonini N, Zoetmulder FA. Survival analysis of pseudomyxoma peritonei patients treated by cytoreductive surgery and hyperthermic intraperitoneal chemotherapy. *Ann Surg*. 2007;245(1):104–109.

36. Chua TC, Quinn LE, Zhao J, Morris DL. Iterative cytoreductive surgery and hyperthermic intraperitoneal chemotherapy for recurrent peritoneal metastases. *J Surg Oncol*. 2013;108(2):81–88.
37. Bijelic L, Yan TD, Sugarbaker PH. Treatment failure following complete cytoreductive surgery and perioperative intraperitoneal chemotherapy for peritoneal dissemination from colorectal or appendiceal mucinous neoplasms. *J Surg Oncol*. 2008;98(4):295–299.
38. Vassos N, Förtsch T, Aladashvili A, Hohenberger W, Croner RS. Repeated cytoreductive surgery (CRS) with hyperthermic intraperitoneal chemotherapy (HIPEC) in patients with recurrent peritoneal carcinomatosis. *World J Surg Oncol*. 2016;14(1):42.
39. Sugarbaker PH, Chang D. Results of treatment of 385 patients with peritoneal surface spread of appendiceal malignancy. *Ann Surg Oncol*. 1999;6(8):727–731.
40. Stewart JH 4th, Shen P, Russell GB, et al. Appendiceal neoplasms with peritoneal dissemination: outcomes after cytoreductive surgery and intraperitoneal hyperthermic chemotherapy. *Ann Surg Oncol*. 2006;13(5):624–634.
41. Yoon W, Alame A, Berri R. Peritoneal Surface Disease Severity Score as a predictor of resectability in the treatment of peritoneal surface malignancies. *Am J Surg*. 2014;207(3):403–407 (**discussion 406–407**).
42. Harmon RL, Sugarbaker PH. Prognostic indicators in peritoneal carcinomatosis from gastrointestinal cancer. *Int Semin Surg Oncol*. 2005;2(1):3.
43. Verwaal VJ, van Ruth S, de Bree E, et al. Randomized trial of cytoreduction and hyperthermic intraperitoneal chemotherapy versus systemic chemotherapy and palliative surgery in patients with peritoneal carcinomatosis of colorectal cancer. *J Clin Oncol*. 2003;21(20):3737–3743.
44. Levine EA, Votanopoulos KI, Shen P, et al. A multicenter randomized trial to evaluate hematologic toxicities after hyperthermic intraperitoneal chemotherapy with oxaliplatin or mitomycin in patients with appendiceal tumors. *J Am Coll Surg*. 2018;226(4):434–443.

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.